

ON THE REGULATION OF THE RESPIRATION IN REPTILES

I. THE EFFECT OF TEMPERATURE AND CO₂ ON THE RESPIRATION OF LIZARDS (*LACERTA*)

By BODIL NIELSEN

The Laboratory of Zoophysiology A, University of Copenhagen

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Ventilation of the lungs accomplished by movements of the ribs is, in the phylogenetic development of the animals, first found in the reptilian class. It may therefore be of interest to investigate the regulation of respiration in this group. The reactions to carbon dioxide and to low oxygen tensions is specially interesting considering the importance of these substances for the respiratory regulation in higher animals and man.

The literature on reptilian respiration is not extensive and is mostly concerned with the respiratory movements in different species: Bert (1870), turtles, snakes, lizards, crocodiles; Langendorff (1891), *Lacerta*, *Anguis*; Siefert (1896), *Lacerta*; Kahn (1902), *Lacerta*; Babak (1914*a, b*), *Iguana*, crocodile; v. Saalfeld (1934*a, b*), *Uromastix*; Willem & Bertrand (1936), *Lacerta*; Vos (1936), turtles, snakes, lizards, crocodiles; and Boelaert (1941), various lacertilians, and (1942), crocodiles.

The chemical regulation of respiration in various species of reptiles has earlier been studied by Siefert (1896), Babak (1914*a, b*), v. Saalfeld (1934*a*), Vos (1936), Boelaert (1941) and Randall, Stullken & Hiestand (1944). A critical survey of the literature was presented by Vos (1936).

The more recent investigations confirm the finding that CO₂ and lack of O₂ have a strong effect on the respiratory pattern of reptiles, but the various studies are often ambiguous and incomplete. Mostly, very high CO₂ and very low O₂ concentrations have been used. The range of concentrations within which a natural regulatory effect of these gases take place is probably much smaller. Investigations of the effect of temperature on respiration are few, and the relation between O₂ uptake and pulmonary ventilation has never been studied.

In the present work, the relationships between temperature, O₂ uptake, and ventilation have been studied; further, the effect of different concentrations of CO₂ in the inspiratory air on respiration has been investigated by a method which gives simultaneous determinations of pulmonary ventilation and oxygen uptake. Two species of lizard (*Lacerta viridis* and *L. sicula*) were used as experimental animals. Their reactions to the respiratory stimuli applied were completely identical.

I. METHODS AND PROCEDURE

The method employed records the changes in volume of the body as a whole, which is the same as a registration of the volume changes in the lungs caused by the pulmonary ventilation. By this method, unanaesthetized animals can be used in repeated

experiments. This is an advantage when the natural respiratory regulation is to be studied.

Apparatus. The experimental set-up is shown in Fig. 1. *A* is a spirometer (volume $\frac{1}{2}$ l. or 1.2 l.) loaded with a weight *l*, so that air is forced out through the resistance *x* through the helmet of the animal container *E* and out past the glass syringes *J*. By changing the weight on the spirometer and the resistance (glass capillaries of different lengths and bores) the airflow through the system can be varied. On the drum *B* the air content in the spirometer *A* is registered and the volume of air forced through the system during the experiment can therefore be measured.

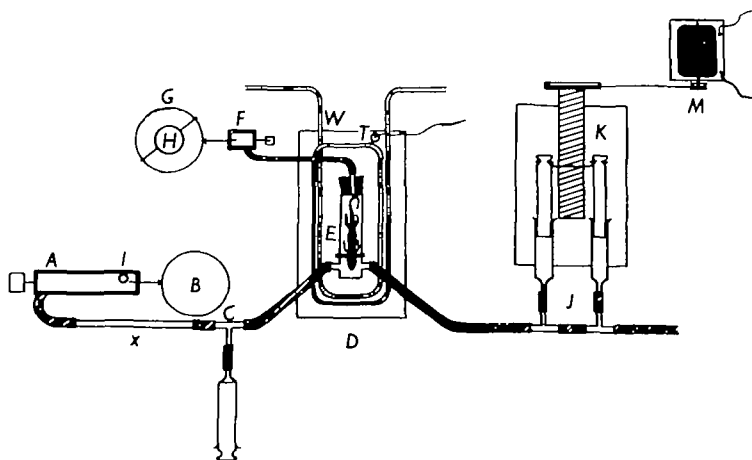


Fig. 1. Apparatus. *A*, spirometer; *B*, drum; *C*, test-tube for air sampling; *D*, water bath; *E*, animal container; *F*, small Krogh-type spirometer for registration of the respirations; *G*, revolving drum; *H*, clockwork; *J*, air-sampling syringes; *K*, sampling device; *M*, electric motor; *T*, contact thermometer of the thermostat; *W*, cooling spiral. *l*, weight; *x*, glass capillary resistance.

C is a test-tube for taking samples of the influx air. *D* is a container filled with water. The temperature within the container can be regulated by leading cold water through the coil *W* or by electrical heating. In the latter case, the thermostat *T* keeps the temperature constant ($\pm \frac{1}{10}^{\circ}$ C.). A small electric motor drives a stirrer and provides an even temperature distribution in the water.

E is the animal container and *F* is a small Krogh-type spirometer (volume 3.5 ml. or 7 ml.) registering the respiratory movements on the drum *G*. *G* is turned at a constant speed (one turn per 30 min., or one per 15 min.) by a clockwork motor *H*.

The electric motor *M* drives the sampling device *K* which slowly pulls back the pistons in the syringes *J*, so that a small part of the air coming from the animal container (the exit air) is continually sampled.

The animal to be used was placed in a separate cage without food and water the day before the experiment. During the experiment, the animal was placed with a tight-fitting rubber diaphragm around its neck, in a container consisting of a body-chamber and a 'helmet'. The helmet and body-chamber, which were firmly screwed together, were thus separated from one another by the air-tight diaphragm.

The animal breathed the air flowing through the helmet, and the volume changes of the body in the body-chamber were transmitted to the spirometer *F* and registered

as a ventilation curve on the revolving drum *G*. The animal container was placed in the water bath *D*, so that the experiment could be performed at constant temperature.

At each experiment the spirometer *A* was filled with the desired air mixture. The flow was so regulated that the difference in CO₂ percentage in the air reaching the helmet (influx air) and leaving the helmet (exit air) was about 0.6%.

When air mixtures different from room air were used, the particular air mixture was bubbled through the water in the spirometer for about 30 min., so that equilibrium between water and air in the spirometer could be established before the experiment. Each experiment lasted 15 min., but readings and sampling were not started until an initial period (15–60 min.) had elapsed, during which the body temperature became constant and the respiration regular.

Immediately before and after the actual experiment, samples of the influx air were taken. The body temperature of the animal could be measured by means of a thermocouple, one junction of which, mounted in a small plastic tube, was inserted 1 cm. into the cloaca of the animal. Usually two experiments were made in succession without touching the animal, which often spent about 2 hr. in the apparatus.

Double analyses for CO₂ and O₂ content of the influx and exit air were made on the Scholander 0.5 ml. analysing apparatus (Scholander, 1947). If the results of a pair differed more than 0.05% on CO₂ or O₂ the experiment was discarded.

Computations. From the volume of the influx air at STPD and the N₂ percentages of the influx and exit air, the volume of the exit air was calculated. Oxygen uptake, carbon dioxide elimination, and R.Q. could then be determined. On the respiration curve, the number of respirations during the experiment was counted and the depth of the respirations was measured. From these data, an average respiratory frequency and an average respiratory depth were estimated. The product of these gave the average pulmonary ventilation per minute during the experiment. (The values were converted to BTPS.)

Accuracy of the method. The accuracy of the results attained by the described method depends on the accuracy of the readings of the volumes of air from spirometer *A* and on the reliability of the analyses. The latter is of special importance when, as here, the difference between influx air and exit air used for computing the metabolism is so small (0.6%).

The error on the volume readings is only about 1%, and the uncertainty of the values for the metabolism can be calculated to be about 10%. The error on the ventilation is of the same order of magnitude, roughly estimated to be 10%, and depends mostly on the readings of the respiratory amplitude.

Even if the values of both metabolism and respiration obtained by this method are encumbered with the above-mentioned uncertainties, they give valuable information. The advantage of giving absolute values for respiratory frequency, depth, and ventilation under different circumstances must be considered great compared to the mere description given in earlier investigations.

II. VENTILATION AND METABOLISM AT DIFFERENT TEMPERATURES

In ten different animals (3 *Lacerta sicula* and 7 *L. viridis*) the oxygen uptake and the pulmonary ventilation were determined as described in Part I.

The experimental conditions were, in all experiments, kept as near to resting conditions as possible. It was very seldom that the animals did not struggle to get free one or more times during the experiment. It must also be mentioned that an animal held (as in the present experiments) in a fixed position is probably not relaxed and has an increased muscle tone which may increase the oxygen uptake above the basal level. However, as these sources of error are present in all the experiments, it is still justifiable to compare them.

The oxygen uptake was varied by changing the body temperature of the animals. It usually took 40–60 min. to cool an animal down from room temperature to 10° C., and about the same time to warm it up to 35° C. Experiments were performed at temperatures of 10°, 20° (room temperature), 30° and 35° C. The relationship between O₂ uptake and temperature is shown in Fig. 2.

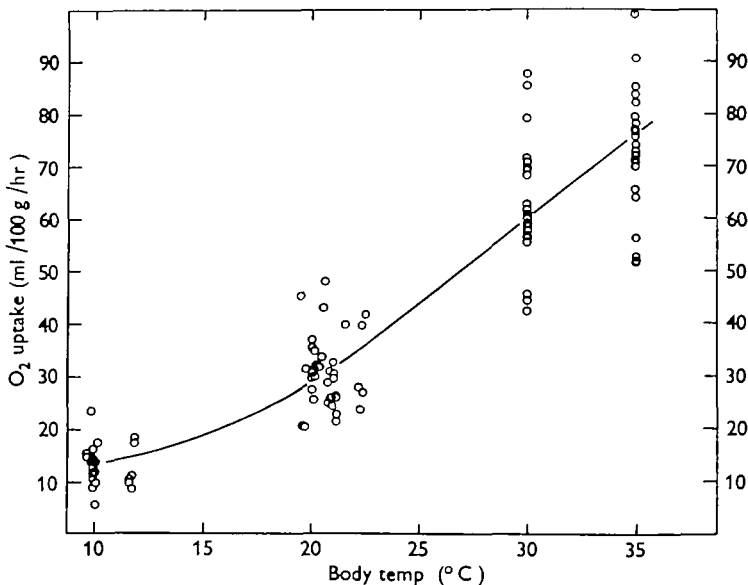


Fig. 2. Oxygen uptake in ml. per 100 g. per hour in relation to body temperature °C. (results from eight animals).

RESULTS

In Fig. 3 the pulmonary ventilation (BTPS) is plotted in relation to the oxygen uptake (STPD). An increase in oxygen uptake gives an increased pulmonary ventilation. The relation is not quite rectilinear.

The oxygen uptake and the ventilation at 10° C. are relatively low. This is probably caused by the lack of struggling at this temperature where the animals are sluggish. In Fig. 4 the respiratory frequency and the amplitude at different temperatures are shown. The respiratory frequency increases with increasing body temperature. The increase seems to be exponential. The depth of the respirations is independent of changes in temperature. In another series, performed on another set of animals, the respiratory depth varied more (as seen in Fig. 5, where the results are given as percentages of the 35° C. value). No relationship, however, seems to exist between

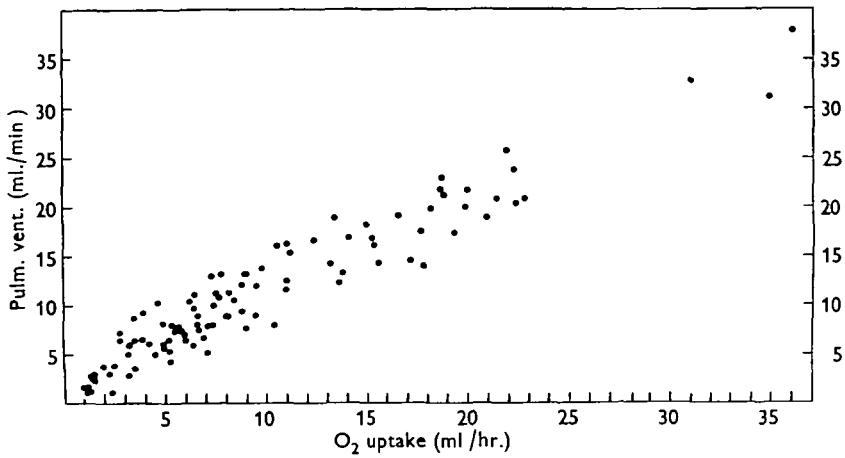


Fig. 3. The relationship between pulmonary ventilation, ml./min. (BTPS) and oxygen uptake, ml./hr. (STPD). The oxygen uptake was varied by changing the body temperature of the animals (results from eight animals combined).

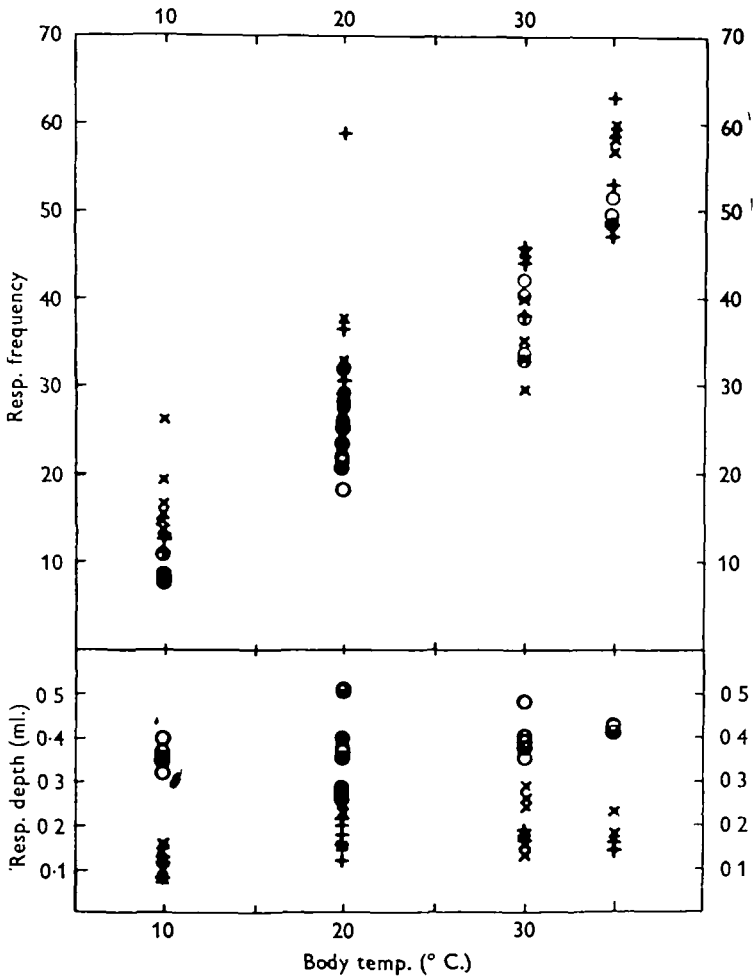


Fig. 4. Above: respiratory frequency in relation to body temperature. Below: respiratory depth in relation to body temperature (four animals: O, ●, + and x).

temperature and respiratory depth. The quotient, *ventilation/hour* ÷ *O₂ uptake/hour* (i.e. the ventilation per litre consumed oxygen), varies a great deal even for the same animal in repeated experiments. Mean values for all animals are shown in Table 1, column 3.

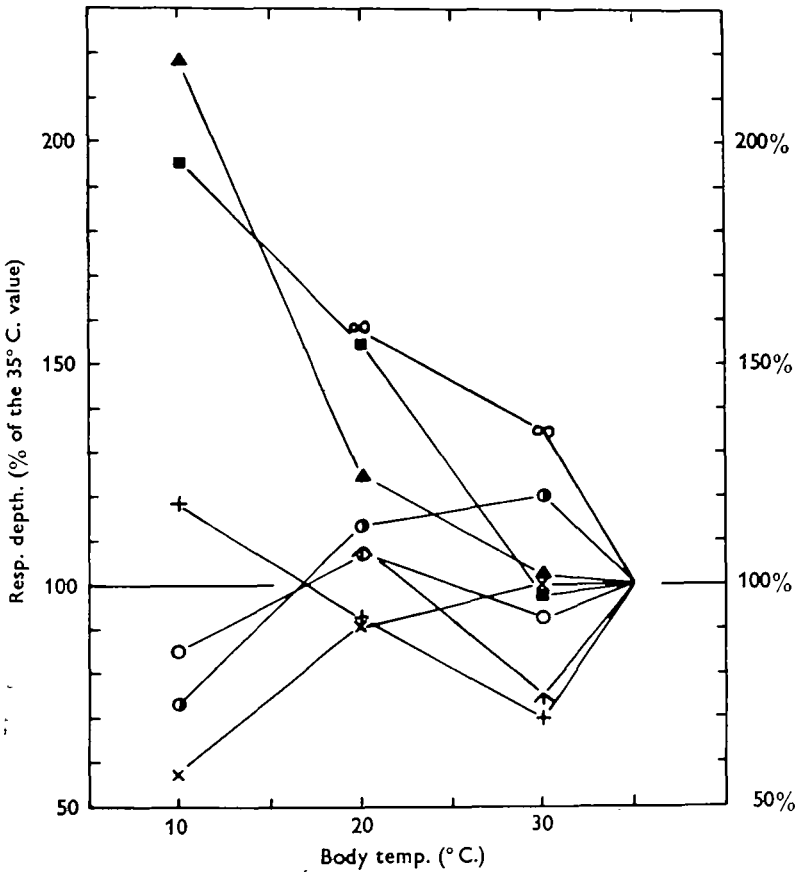


Fig. 5. Respiratory depth in percentage of the 35° C. value in relation to body temperature (eight animals: O, ×, ●, +, ▲, ■, ∞ and ↑).

Table 1

Temp. (°C.)	No. of experiments <i>n</i>	Vent./hr. (BTPS)	Standard error of the mean <i>s</i>	Vent./hr. (STPD)	R.Q. mean	Vent./hr. (STPD)
		O ₂ uptake/hr. (STPD), mean <i>m</i>		O ₂ uptake/hr. (STPD), mean <i>m</i> ₁		CO ₂ output/hr. (STPD), mean
ca. 10	22	96.8	± 7.89	92.1	0.88	104.7
ca. 20	38	87.2	± 3.81	79.2	0.80	99.0
30	23	65.4	± 2.93	56.0	0.81	69.1
35	23	66.0	± 2.05	55.2	0.82	67.3

The differences between the ventilation quotients at 10° and 30° C., 10° and 35° C., 20° and 30° C., and at 20° and 35° C. are highly significant, while the differences between the quotients at 10° and 20° C., and 30° and 35° C. are not statistically significant.

The high numerical value of the quotient (65-97, as compared to 20-24 in man) is probably due to the rather primitive structure of the lungs (cf. Milani, 1894).

DISCUSSION

This part of the investigation showed that the rise in metabolic rate caused by an increase in temperature is accompanied by an increase in pulmonary ventilation. The increase is brought about by an increase in respiratory frequency, whereas the respiratory depth remains practically unchanged.

The increased ventilation may be caused by an increased blood $p\text{CO}_2$ or by the increased temperature *per se*, or, perhaps, by a combination of both factors. An increased blood $p\text{CO}_2$ could be caused simply by the increased production of CO_2 at the higher temperatures. No values of blood $p\text{CO}_2$ are available, but certain conclusions as to its variations may be drawn from the experimental results.

The CO_2 output per minute can be expressed as:

$$\text{CO}_2 \text{ production} = \text{pulmonary ventilation (STPD)} \\ \times (\text{expired CO}_2 \% - \text{inspired CO}_2 \%),$$

from which follows that

$$\frac{\text{pulm. vent. (STPD)}}{\text{CO}_2 \text{ production (STPD)}} = \frac{1}{(\text{exp. CO}_2 \% - \text{insp. CO}_2 \%)}$$

The left-hand expression decreases as temperature increases (Table 1, column 7). Consequently, $(\text{exp. CO}_2 \% - \text{insp. CO}_2 \%)$ must be increasing. As the inspired CO_2 percentage is kept nearly constant (it must be equal to the average CO_2 percentage in the helmet, which again is approximately equal to that in the exit air), it follows that the CO_2 percentage in the expired air must be higher at the higher temperatures. With a constant respiratory depth (and a presumably constant dead space) it can be concluded that the alveolar CO_2 percentage and hence the blood and tissue $p\text{CO}_2$ is also higher at higher temperatures. The increased ventilation must, therefore, at least partly be caused by an increased CO_2 stimulus on the respiratory centre.

This effect of the increased tissue and blood $p\text{CO}_2$ on respiration is apparently different from the effect of CO_2 added to the inspired air in that it increases the respiratory frequency and ventilation, whereas inspired CO_2 increases the respiratory depth and slows the frequency as will be discussed in Part III.

An effect of increased temperature alone on respiration has been demonstrated by v. Saalfeld (1934*a*). He found that local heating of a leg of *Uromastix*, from which the skin was removed and all nerve connexions cut, was followed by an increased ventilation. This effect could be prevented if the neck of the animal (carotid and vertebral arteries) was cooled. From this he concluded that the rise in pulmonary ventilation was caused by a heating of the respiratory centre by the blood. He assumed that, by a general heating of the animal, metabolites from the increased metabolism should further act as stimuli for the respiratory centre, as cooling in this case did not completely abolish the ventilatory increase.

In humans, Cunningham & O'Riordan (1957) have found that a raised temperature increases the sensitivity of the respiratory centre towards CO_2 . An interaction of this kind between temperature and CO_2 might, in the case of *Lacerta* also, be one of the

reasons for the increase in pulmonary ventilation at higher temperatures. Finally, metabolic factors other than CO_2 could be involved in the ventilatory response to increased temperature. Whether the increase is caused by one or several of the above-mentioned factors cannot be decided at present. Determinations of $p\text{CO}_2$ in blood and alveolar air may give interesting results.

In the temperature interval studied here the rise in pulmonary ventilation is not connected with temperature regulation. The rise is called forth by the metabolic requirements. This can be seen from the ratio *pulmonary ventilation/litre of O_2 consumed*. This quotient is not higher at higher temperatures as it would have been if the ventilatory rise was due to thermal panting. On the contrary, the value of *ventilation (BTPS)/hr. \div O_2 uptake (STPD)/hr.* at both 30° and 35° C. is smaller than at 10° and 20° C. At still higher temperatures it is quite possible that *Lacerta* also would show thermal panting [cf. Langlois (1901), Cowles & Bogert (1944), and others].

The better utilization of the respiratory air found at higher temperatures (cf. *ventilation (STPD)/ O_2 uptake (STPD)*, Table 1) may be due to a larger, and perhaps better distributed, blood flow through the lungs. It cannot, however, be due to a relatively decreased anatomical dead space, as the respiratory depth is not greater but remains unchanged or even becomes smaller at 30° and 35° C.

III. PULMONARY VENTILATION AND OXYGEN UPTAKE AT DIFFERENT CO_2 PERCENTAGES

Five lizards (2 *Lacerta sicula* and 3 *L. viridis*) were used for the study of the influence of CO_2 on respiration. The procedure is described in Part I. Pulmonary ventilation, respiratory frequency and depth, and oxygen uptake were measured.

The composition of the actually inspired or expired air is not known and cannot be computed from the values measured in the experiments. The exit air, however, must fairly accurately represent the average composition of the air in the helmet from which the animal inspires. By changing the flow rate and the composition of the influx air, the exit air can be maintained at a relatively constant composition for any CO_2 percentage desired.

The CO_2 percentage of the expired air and the alveolar air is naturally higher than that of the exit (and influx) air. At a constant CO_2 production (i.e. rest at constant temperature) an increase in CO_2 percentage in the helmet (exit air) would produce an equal increase in the expired air if the ventilation did not change. If, however, the ventilation increases, the increase in CO_2 percentage of the expired air will be less than that of the exit air.

In the following paragraphs the changes in pulmonary ventilation and respiratory pattern will be related to the CO_2 percentage in the exit air. It must be understood that the changes in alveolar CO_2 percentages may be smaller than those of the exit air, i.e. in cases where the alveolar ventilation has increased.

RESULTS

When the CO_2 percentage of the exit air is increased, the respiratory pattern changes. CO_2 percentages below 3% in the exit air causes a gradual increase in respiratory depth and pulmonary ventilation and a decrease in respiratory frequency

When the CO_2 percentage is increased above 3–4% it produces a 'periodic inhibition' of respiration. Groups of respirations separated by inspiratory pauses lasting up to 1 min. occur. The duration of this periodic inhibition ('Cheyne-Stokes-like' respiration) is dependent on the CO_2 percentage, lasting from a few minutes at 3% to more than 1 hr. at 13.6%. In this state the respiratory depth is increasing, while the pulmonary ventilation naturally is very low due to the disturbed breathing rate. This is illustrated by Figs. 6*a*, 7 and 8, where the effect of 7.2% CO_2 is shown.

After 20–60 min. respiration always becomes adjusted to the CO_2 percentage and is unchanged and regular from then on (steady state, Fig. 6*b*).

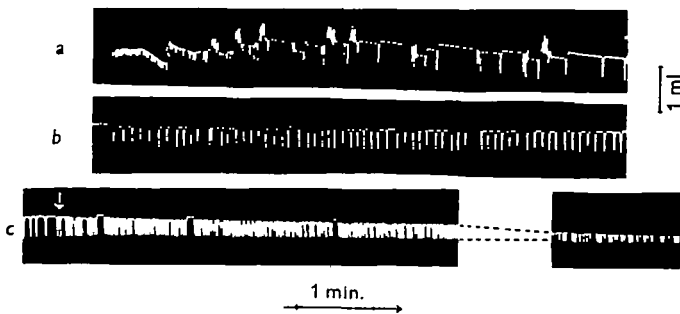


Fig. 6. Respiration curves: (a) Start of CO_2 breathing, note Cheyne-Stokes respiration and long inspiratory pauses. (b) Steady state, 7.2% CO_2 in exit air. (c) Change from CO_2 breathing to room air breathing at ↓ (read left to right).

After a steady state has been reached, a sudden shift back to room air with quick flow causes an instantaneous increase in respiratory frequency and, consequently, also in pulmonary ventilation (cf. Figs. 7 and 8). The ventilations attained here are the highest registered for the animals concerned. The respiratory frequency then decreases gradually, reaching the normal steady-state value after 2–10 min.

During the same time the respiratory depth after the shift to room air decreases slowly and, consequently, the pulmonary ventilation also decreases. Normal values for respiratory depth and ventilation are reached after 20–30 min. (Figs. 7, 8).

In Fig. 9 the pulmonary ventilation in the steady state is plotted in relation to the CO_2 percentage in the exit air. It is seen that an increase in CO_2 to about 3% causes a slight increase in pulmonary ventilation. At further increases in the CO_2 percentage the ventilation again decreases, even to subnormal values. This maximum, at about 2.75% CO_2 , was observed in all five animals investigated. In the most CO_2 -sensitive animal the ventilation was doubled at this CO_2 concentration.

Fig. 9 shows that in the steady state the respiratory depth increases regularly with increasing CO_2 percentage in the exit air (0.4–13.6%), while the respiratory frequency decreases. These relationships between CO_2 and respiratory depth and frequency was found to be unaffected by a change in the animal's body temperature from 20° to 30° C. The shape of the curves obtained at 30° C. was similar to that of the 20° C. curves, the frequency curve lying higher, and the respiratory depth curve a little below the corresponding 20° C. curves.

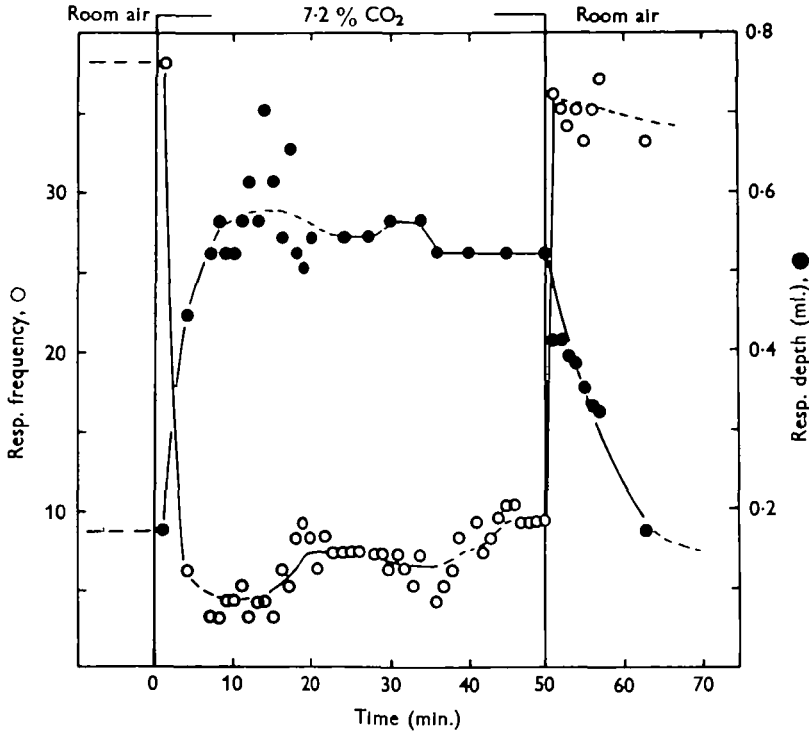


Fig. 7. Respiratory changes produced by changing from room air breathing to CO_2 breathing (50 min.) and back to room air. CO_2 percentage 7.2 in exit air. Time in minutes. O, Respiratory frequency; ●, respiratory depth (one animal). Temp. 20°C .

DISCUSSION

The results presented in Figs. 6–9 show that CO_2 in the inspired air influences respiration markedly at all percentages used. At percentages lower than 3% CO_2 causes an increased pulmonary ventilation, while higher percentages cause a decrease in pulmonary ventilation to subnormal values. Higher CO_2 percentages (above 3–4%) cause further transitory disturbances in the respiratory pattern ('periodic inhibition') by causing a 'Cheyne-Stokes-like' respiration.

Babak (1914a, b), v. Saalfeld (1934a), Vos (1936), and Boelaert (1941) found 'dyspnoea' at low (< 5% CO_2) and 'inhibition' at higher CO_2 percentages. But most of their experiments on the influence of CO_2 on respiration lie outside the interval where CO_2 stimulates the *pulmonary ventilation*. Siefert (1896) used 100% CO_2 ; the narcotic effect therefore was dominant in his experiments. Randall *et al.* (1944) found that, after a period of apnoea, respiration was stimulated by CO_2 even in very high percentages. This may be due to the fact that their CO_2 experiments lasted only until the appearance of the first groups of vigorous respirations after the apnoeic period (at the most 8 min.). Thereafter they shifted to room air again. This could, perhaps, give the false impression that the pulmonary ventilation is high even in the early period of CO_2 breathing where the respiratory frequency is reduced.

The present study shows that an increase in the CO_2 percentage in the inspired air causes an increase in respiratory depth (Fig. 9). The gradual increase in respiratory depth after the beginning of CO_2 breathing (Fig. 7) may imply that the respiratory depth follows the $p\text{CO}_2$ changes of the blood. It is presumed that the $p\text{CO}_2$ increases gradually from the beginning of the CO_2 breathing and, after some time, reaches a steady level. On shifting from CO_2 breathing back to room air breathing, the respiratory depth again decreases slowly (as opposed to the immediate change in frequency). This might also correspond to the presumably slow fall in the $p\text{CO}_2$ of the blood caused by the gradual CO_2 release from the tissues on returning to room air breathing. However, the respiratory depth did not increase when the $p\text{CO}_2$ of the blood was increased by raising the temperature (metabolic rate) of the animal. It is also possible that the increased respiratory depth may have been influenced by an oxygen lack in the respiratory centre, this oxygen lack being produced by the long apnoeic pauses after the start of the CO_2 breathing and maintained by the low respiratory rate in the steady state. This assumption, however, needs special investigation.

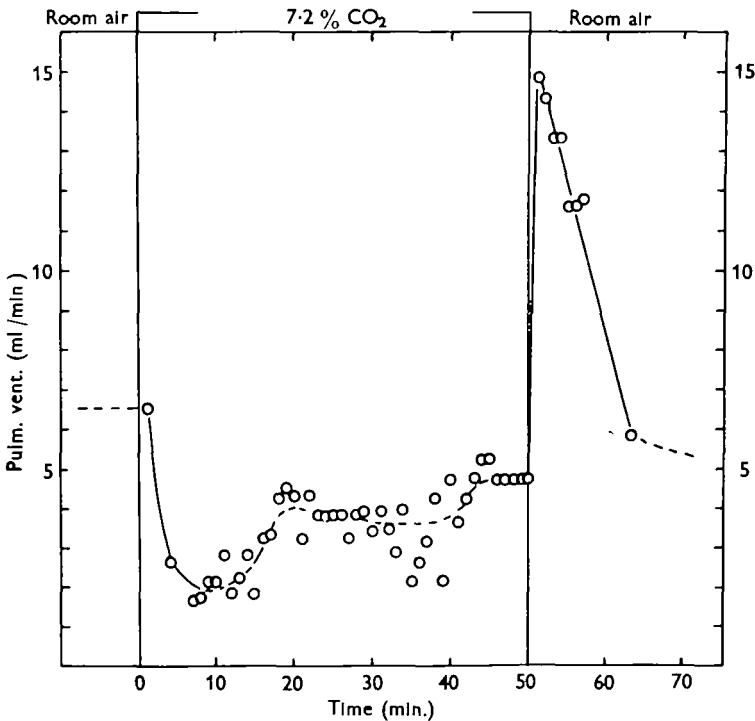


Fig. 8. As Fig. 7 (see Fig. 7), but showing the variation in pulmonary ventilation.

As for the respiratory frequency, no simple correlation seems to exist between frequency and blood $p\text{CO}_2$. At the beginning of CO_2 breathing and in the steady state, the respiratory frequency is the lower the higher the $p\text{CO}_2$ is in the blood.

An increase in $p\text{CO}_2$ produced by an increase in metabolic rate, however, increases the respiratory frequency (see discussion in Part II). It seems, then, that an increased blood and tissue $p\text{CO}_2$, produced by an increase in CO_2 content in the inspired air,

causes the frequency to decrease, while an increased CO_2 tension in the tissues, when the CO_2 content in the inspired air is low, causes the frequency to increase.

The steady-state relationship between frequency and CO_2 content in the inspired air can be explained by the assumption that the respiratory frequency is depressed via chemoreceptive nerve endings in the lungs. An increase in activity of these

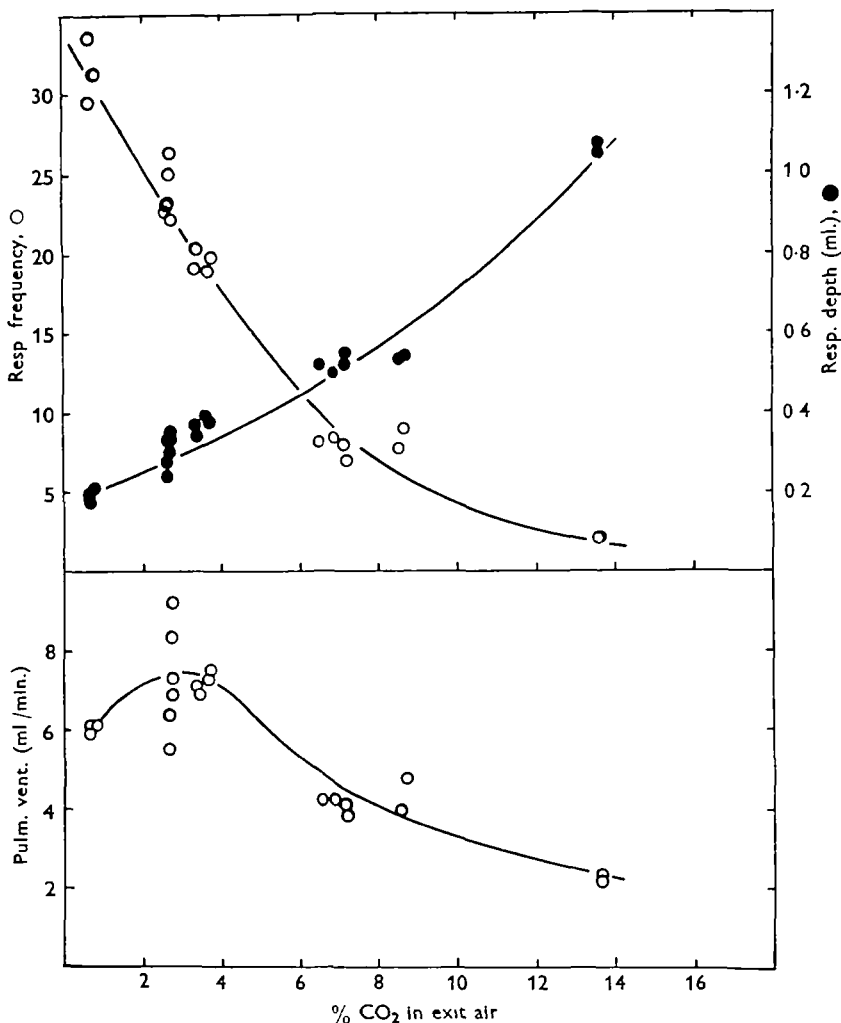


Fig. 9. Respiratory frequency, depth, and pulmonary ventilation in relation to the CO_2 percentage in the exit air. Above: O, respiratory frequency; ●, respiratory depth. Below: O, pulmonary ventilation (one animal). Temp. 20°C .

chemoreceptors might occur as a response to increasing CO_2 content in the lungs. Experiments of v. Saalfeld, Vos and Boelaert have shown that such chemoreceptors exist and that the receptors must be situated in the lungs, not in the upper respiratory pathways. Thus the 'periodic inhibition' at the beginning of CO_2 breathing with higher CO_2 percentage is thought to be a reflex (Babak, v. Saalfeld, Vos and Boelaert). Boelaert (1941) concluded that inhibitory impulses from the chemoreceptors in the

lungs reach the respiratory centre via the vagus nerve, as he found that the inhibition (depression of the frequency) was abolished when the vagus was cut. The causes of the return of the 'Cheyne-Stokes' respiration during prolonged CO₂ breathing to a regular pattern may be thought to be due to an adaptation of the postulated pulmonary chemoreceptors to the CO₂ stimulus, whereas the sudden increase in respiratory frequency, on shifting from CO₂ breathing to room air breathing, would then be due to the disappearance of the depression as room air enters the lungs. The respiratory frequency in the first few minutes after the shift is *higher than the normal* (about twice the normal steady state frequency). This may be an effect of the still high blood and tissue pCO₂ on the respiratory centre. As in the experiments with increased body temperature the CO₂ content in the inspired air is now low and an increased tissue pCO₂ seems to increase the respiratory frequency as shown in Part II.

In the steady state of CO₂ breathing, the inhibitory effect of CO₂ via the lung receptors may veil this direct accelerating effect that blood and tissue CO₂ seems to have on the frequency. However, no correlation between the previous CO₂ percentage and the maximum value of respiratory frequency after the shift to room air was found in the present experiments.

As for the pulmonary ventilation, this study seems to show that in *Lacerta* the pulmonary ventilation is not primarily regulated by CO₂. In the steady state the ventilation could, at the most, only be doubled by CO₂ administration. This small ventilatory increase (the result of the CO₂ effect on the respiratory frequency) is especially striking when the great ability of the animals to increase both respiratory frequency and depth is considered. According to Babak, a combination of low O₂ concentration and CO₂ in the inspired air gives a heavy 'dyspnœe' in *Iguana*. In man, Nielsen & Smith (1951) similarly have shown that, during hypoxia, the effect of CO₂ on the pulmonary ventilation is much increased. It seems possible that in the case of *Lacerta* also, a combination of CO₂ and low oxygen percentages can stimulate the pulmonary ventilation to much higher values than CO₂ administration alone.

SUMMARY

1. In two species of *Lacerta* (*L. viridis* and *L. sicula*) the effects on respiration of body temperature (changes in metabolic rate) and of CO₂ added to the inspired air were studied.

2. Pulmonary ventilation increases when body temperature increases. The increase is brought about by an increase in respiratory frequency. No relationship is found between respiratory depth and temperature.

3. The rise in ventilation is provoked by the needs of metabolism and is not established for temperature regulating purposes (in the temperature interval 10°–35° C.).

4. The ventilation per litre O₂ consumed has a high numerical value (about 75, compared to about 20 in man). It varies with the body temperature and demonstrates that the inspired air is better utilized at the higher temperatures.

5. Pulmonary ventilation increases with increasing CO₂ percentages in the inspired air between 0 and 3%. At further increases in the CO₂ percentage (3–13.5%) it decreases again.

6. At each CO₂ percentage the pulmonary ventilation reaches a steady state after some time (10–60 min.) and is then unchanged over prolonged periods (1 hr.).

7. The respiratory frequency in the steady state decreases with increasing CO₂ percentages. The respiratory depth in the steady state increases with increasing CO₂ percentages. This effect of CO₂ breathing is not influenced by a change in body temperature from 20° to 30° C.

8. Respiration is periodically inhibited by CO₂ percentages above 4%. This inhibition, causing a Cheyne–Stokes-like respiration, ceases after a certain time, proportional to the CO₂ percentage (1 hr. at 8–13% CO₂), and respiration becomes regular (steady state). Shift to room air breathing causes an instantaneous increase in frequency to well above the normal value followed by a gradual decrease to normal values.

9. The nature of the CO₂ effect on respiratory frequency and respiratory depth is discussed, considering both chemoreceptor and humoral mechanisms.

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